Assessment of Effect of Different Packaging Materials on Microbial Quality of Locust Beans during Storage

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Authors’ contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AFSJ/2023/v22i849

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/100350

Received: 22/03/2023
Accepted: 24/05/2023
Published: 28/06/2023

Original Research Article

ABSTRACT

This study assesses the effect of different packaging materials on the microbial quality of Africa locust beans during storage. Locust beans (Dawadawa) also known as iru, among the Yorubas in South-west Nigeria, is a popular condiment used as a taste and flavour enhancer in soup and dishes in Africa. Locust bean is traditionally produced from locust beans seed (Parkia biglobosa) and preserved using different packaging materials before use in order to prolong its shelf life. The Microbiological examination of the produced locust beans was carried out on samples wrapped with different packaging materials such as plastic containers, nylon, and dry banana leaves. The suspected organisms isolated include Salmonella spp, Staphylococcus aureus, Streptococcus lactis, Pseudomonas aeruginosa, Lactobacillus plantarium, Bacillus cereus, Bacillus subtilis, Leuconostoc sp, Escherichia coli and Micrococcus sp. It was observed from this study that locust beans packaged with banana leaves had the highest microbial load, few of which are of public health concern while plastic containers had the least microbial load. It is therefore recommended that locust beans should be produced and packaged with sealed plastic containers to avoid contamination by pathogenic organisms.

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Keywords: Locust beans; packaging; pathogenic organisms.

1. INTRODUCTION

“African locust beans (Parkia biglobosa) also known as iru, among the Yorubas in South-west Nigeria, are popular condiment used as a taste and flavour enhancer in soup and dishes in Africa. Dawadawa has traditionally produced from locust beans (Parkia biglobosa) seeds” [1]. Stated that “fermented locust bean seed is commonly consumed in Ghana, Nigeria, Sierra Leone, and Togo [2]. In Nigeria it is called iku in Yoruba, dawadawa in Hausa and ogiri “igala in igbo”. It is also referred to as kinda in Sierra Leone and kpalugu in Ghana. Preservation and preservatives are designed to inhibit/control the activities of spoilage - causing organisms in food, a process also referred to as sanitization” [2]. “African locust bean is a nutritious source of food since it is rich in protein and some beneficial health components. It serves as a cheap source of protein for people whose source of protein is low due to the high cost of animal protein sources” [3].

“Food packaging is an integral part of food processing, which entails the use of some materials in the wrapping of foods” [4]. “The success of most preservation methods depends on appropriate packaging. However, faulty packaging will undo all that a good processor has attempted to accomplish through the most meticulous manufacturing process” [5].

“Spoilage - causing organisms due to their growth, and metabolic activities produce by-products, which change the texture, taste, flavor and the aroma of the food” [3]. “The use of packaging and preservation of locust beans is to make the product shelf life stay fresher and longer. Dehydration, salting, and packaging of home-produced food items such as locust beans (iru) in simple polyethylene bags are desirable to extend their shelf life” [1].

“In Nigeria, the African locust bean (Parkia biglobosa) tree grows widely throughout the savanna” [6]. “A mature pod contains yellow, dry, and powdery pulp in which dark brown seeds are embedded. The pulp is licked for its sweet taste but only to a small extent. The pulp is usually washed away when the seeds are processed into a condiment called dawadawa or iku. Dawadawa is a source of protein intake among the low-income groups and rural populations of West Africa” [6]. While the seed has been extensively studied [7], little has been done on the utilization of the pulp. “In West Africa, the pulp is prepared as flour and used in soups and stews or eaten with cereals as porridge” [8].

“Plants are known to contain high amounts of essential nutrients, vitamins, minerals, fatty acids and fibre” [9]. “Plants also contain other chemical compounds such as saponins, tannins, oxalates, phytales, trypsin inhibitors, and cyanogenic glycosides, which are known as secondary metabolites and are biologically active” [10]. “The seeds on fermentation are used in cooking stew and soup. It has been reported that husks and pods are good for livestock” [11]. “The roots, barks, leaves, stems, flowers, fruits, and seeds of P. biglobosa are all used medicinally to treat a range of ailments, including diarrhea ulcers, pneumonia, burns, coughs, etc” [12].

Different packaging materials are used for wrapping locust beans to extend their shelf life and the microbial load of these packaging materials is mostly not assessed due to ignorance and little information on its effect on the product. It is therefore essential to assess the effect of different packaging materials used for the storage and preservation of the locust beans. This will ascertain the best packaging materials with the lowest microbial load suitable for their preservation. This will give information on how to improve the shelf life of locust beans and encourage its acceptability.

2. MATERIALS AND METHODS

2.1 Sample Collection and Preparation

Parkia biglobosa (Africa locust bean) seeds used for this research were purchased from Oja-Oba market in Ado-Ekiti, Ekiti-State, Nigeria.

2.2 Sample Preparation

The production and fermentation process was carried out in the Department of Food Technology Laboratory, School of science and computer studies. Federal polytechnic Ado-Ekiti, Ekiti-State Nigeria

2.3 Production of Fermented Parkia biglobosa (Africa locust bean) “iru” using Traditional Method

Parkia biglobosa (Africa locust bean) seeds was prepared according to the method of [13]. Parkia biglobosa (Africa locust bean) seed were sorted and washed to remove the yellowish pulp from the seeds.
### Table 1. Biochemical characteristics of isolates from African locust beans package with plastic container

<table>
<thead>
<tr>
<th>Isolates</th>
<th>Grams Rxn</th>
<th>Shape</th>
<th>Catalase</th>
<th>Indole</th>
<th>Vp</th>
<th>Oxidase</th>
<th>Methyl red</th>
<th>Citrate</th>
<th>Probable organisms</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>-</td>
<td>rod</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>Salmonella spp</td>
</tr>
<tr>
<td>2</td>
<td>+</td>
<td>Cocci in cluster</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Staphylococcus aureus</td>
</tr>
<tr>
<td>3</td>
<td>-</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Pseudomonas aeruginosa</td>
</tr>
<tr>
<td>4</td>
<td>+</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>Bacillus cereus</td>
</tr>
<tr>
<td>5</td>
<td>+</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>Bacillus subtilis</td>
</tr>
</tbody>
</table>

**Key:**
- + means positive
- - Means negative

### Table 2. Biochemical characteristics of isolates from African locust beans packaged with nylon

<table>
<thead>
<tr>
<th>Isolates</th>
<th>Grams Rxn</th>
<th>Shape</th>
<th>Catalase</th>
<th>Indole</th>
<th>Vp</th>
<th>Oxidase</th>
<th>Methyl red</th>
<th>Citrate</th>
<th>Probable organisms</th>
</tr>
</thead>
<tbody>
<tr>
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<td>rod</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>Salmonella spp</td>
</tr>
<tr>
<td>2</td>
<td>+</td>
<td>Cocci in chains</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>Streptococcus Lactis</td>
</tr>
<tr>
<td>3</td>
<td>-</td>
<td>Bacilli</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Pseudomonas aeruginosa</td>
</tr>
<tr>
<td>4</td>
<td>+</td>
<td>Bacilli</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>Lactobacillus plantarium</td>
</tr>
<tr>
<td>5</td>
<td>+</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>Bacillus cereus</td>
</tr>
<tr>
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<td>+</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>Bacillus subtilis</td>
</tr>
<tr>
<td>7</td>
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<td>coccobacilli</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Leuconostoc sp</td>
</tr>
<tr>
<td>8</td>
<td>+</td>
<td>Spherical</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>Micrococcus sp</td>
</tr>
</tbody>
</table>

**Key:**
- + means positive
- - Means negative
Table 3. Biochemical characteristics of isolates from African locust beans from leaves

<table>
<thead>
<tr>
<th>Isolates</th>
<th>Grams Rxn</th>
<th>Shape</th>
<th>Catalase</th>
<th>Indole</th>
<th>Vp</th>
<th>Oxidase</th>
<th>Methyl red</th>
<th>Citrate</th>
<th>Probable organisms</th>
</tr>
</thead>
<tbody>
<tr>
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<td>rod</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>Salmonella spp</td>
</tr>
<tr>
<td>2</td>
<td>+</td>
<td>Cocci in cluster</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Staphylococcus aureus</td>
</tr>
<tr>
<td>3</td>
<td>+</td>
<td>Cocci in chains</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>Streptococcus lactis</td>
</tr>
<tr>
<td>4</td>
<td>-</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>Pseudomonas aeruginosa</td>
</tr>
<tr>
<td>5</td>
<td>+</td>
<td>Bacilli</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Lactobacillus plantarium</td>
</tr>
<tr>
<td>6</td>
<td>+</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>Bacillus cereus</td>
</tr>
<tr>
<td>7</td>
<td>+</td>
<td>rod</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>Bacillus subtilis</td>
</tr>
<tr>
<td>8</td>
<td>-</td>
<td>coccobacilli</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Leuconostoc sp</td>
</tr>
<tr>
<td>9</td>
<td>-</td>
<td>rod</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>Escherichia coli</td>
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<tr>
<td>10</td>
<td>+</td>
<td>Spherical</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>Micrococcus sp</td>
</tr>
</tbody>
</table>

Key:
+ means positive
- Means negative
The locust bean seed was cooked with a pressure pot. Put on electric cooker for 7 hours, with water being added frequently to it, until the seed becomes softer, then the seed was allowed to cool down and it was pounded by using mortar and pestle, the cotyledon was separated from its coat. Inside a bowl of water eliminated by using a hand and sieve then the seed was washed and cleaned thoroughly. It was then re-cooked for another 1 hour 30 minutes with pressure pot, until the seed becomes softer. Then the water was drained off by using a sieve. The locust bean was spread into a big sieve wrapped with multiple cloth to maintain a hot atmosphere to be fermented for 72 hours. After fermentation the locust bean “iru” was packaged in three different materials (Banana leaf, Plastic rubber and nylon) and preserved for 7 days.

2.4 Media Preparation

The media (Nutrient agar, MacConkey agar, Salmonella shigella agar and Eosin Methylene blue agar) used for this research were prepared according to the manufacturer’s instructions. The prepared media were poured into conical flasks and the flasks were covered with cotton wool and Aluminum foil. The media were sterilized and autoclaved at 121°C for 15 minutes and used for microbial analysis [14].

2.5 Microbiological Analysis of Fermented Sample

One gram of the sample (Fermented Parkia biglobosa (Africans locust bean) was taken respectively and marched using mortar and pestle for proper homogenization. One gram of sample was diluted serially in seven-fold dilution blanks and properly mixed with sterile glass rod. 0.1ml of the sample was introduced into sterile plates and agar. The media was poured at 45°C. The plates were rotated gently to disperse the inoculum in the medium and allowed to solidify. Then the plates were incubated at 37°C and the resulting colonies of microbes were sub-cultured to get the colonies in their pure form and to lower the density of cells. The culture was maintained as slants on both nutrient Agars. The slants were kept and stored for further use.

2.6 Examination of Culture Plates of Isolated

Macroscopic: the colonial appearance of the organism was noted such as color, shape, and size after 24 hours of incubation.

2.7 Characterization and Identification of Sample

The Gram Staining reaction was carried out by emulsifying one isolate 24 hours old colony in a drop of water placed at the center of a clean grease tree slide passed through a Bunsen burner. The Heat - fixed smear was flooded with crystal violet for 60 seconds, after which the stain was poured off the slide and rinsed with running tap water. The smear was flooded with iodine.

2.8 Identification of Bacteria Isolates

The bacterial isolates were identified by using morphological and biochemical tests such as coagulase test, catalase test, oxidase test, indole test, urea production, nitrate reductions, motility test, spore test, hydrogen sulfide production test, oxidase production, citrate utilization test, nitrate reduction test, Voges Proskauer and Methyl red test [15] and [16].

3. RESULTS

The results of the biochemical characterization of the microorganisms isolated from locust beans wrapped with different packaging materials such as plastic container, nylon and dry banana leaves are shown in the tables below.

4. DISCUSSION

The suspected isolates recorded in Table 1 includes Salmonella spp, Staphylococcus aureus, Pseudomonas aeruginosa, Bacillus cereus and Bacillus subtilis. Salmonella species have been reported to be transmitted through contaminated water and this bacterium is a suspected carrier found in food handlers [17]. Bacillus species were the predominant microorganisms in this sample. The result of this study justifies the assertion of [18] that Bacillus species was the predominant bacteria involved in the fermentation of ogiri which is also a fermented soup condiment. Pseudomonas isolated in this sample is an aerobic organism and thrives well in the presence of oxygen and water [19]. The locust bean seed usually comes with a substantial microbial local comprising spore of aerobic spore-forming bacterial.

The result on Table 2 shows the characteristics of isolates from those packaged with nylon. Suspected organisms include; Salmonella spp
Streptococcus lactis, Pseudomonas aeruginosa, Lactobacillus plantarum, Bacillus cereus, Bacillus subtilis, Leuconostoc sp, and Micrococcus sp. Eight isolates were isolated from the sample. Bacillus spp. which is a predominant organism obtained in this study and Leuconostoc spp. are capable of increasing the protein and fat contents of locust beans. This is similar to the study of [20] who observed that various Bacillus species were responsible for the fermentation of African Locust Bean seeds. The result obtained from this study is similar to the work of [5] who also isolated Lactobacillus, Pseudomonas and bacillus from fermented melon seed.

The result from Table 3 shows the biochemical characteristics of isolates from African Locust beans packaged with dry banana leaves. The suspected isolates are Salmonella spp, Staphylococcus aureus, Streptococcus lactis, Pseudomonas aeruginosa, Lactobacillus planetarium, Bacillus cereus, Bacillus subtilis, Leuconostoc sp, Escherichia coli and Micrococcus sp. The microorganisms isolated from locust beans packaged with dry banana leaves might be due to the contamination from leaves used as packaging materials. The leaves are usually picked up from the ground and on the farm land and are not normally washed thereby introducing different microorganisms from soil and insect droppings. Studies by [21] and [2] has shown that “several microorganisms are associated with locust bean fermentation. He also noted that the most abundant and the major dominant agent of fermentation after 24 hours was Bacillus subtilis. They also noted the presence of Leuconostoc mesenteroides and Staphylococcus species after fermentation”. Similar report also showed that “Bacillus subtilis, Leuconostoc mesenteroides and Staphylococcus sp. were associated with the fermentation of African locust bean seeds to produce “iru” condiment” [8]. However, the presence of Escherichia coli and Staphylococcus aureus are of public health concern and can lead to foodborne diseases. The microbes may be introduced into the products through the contaminated hands of handlers, unsterilized utensils, other materials being used for the processing and even from the air from the environment during the preparation of the product [22,23,24].

5. CONCLUSION

The results from this study show that the use of plastic container as a packaging material should be encouraged when compare to nylon and leaves for packaging of locust beans before consumption. Locust beans packaged with dry banana leaves harbor a lot of microorganisms that can cause foodborne diseases which are therefore of public health concern. It is therefore recommended that locust beans produced must be well preserved with the best packaging materials in order to prevent the entry and contamination by microorganisms.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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